

STANDARD OPERATING PROCEDURES
DIVISION OF COMPARATIVE MEDICINE
UNIVERSITY OF SOUTH FLORIDA

SOP# 418.4

IV. ANESTHESIA AND ANALGESIA

1. General anesthesia is provided using a suitable inhalant anesthetic agent (e.g., isoflurane) with appropriate scavenging of the waste anesthetic gas.
2. Analgesia is provided using a suitable systemic anesthetic agent (e.g., carprofen, 5-10 mg/kg SQ).

V. TISSUE HARVEST

1. Prior to collecting tissue(s) for a PI ensure that procedures for handling the collected samples and methods of identifying both samples and animals are clearly understood.
2. Anesthetize the mouse if required, or gently but securely restrain the mouse.
3. Swab the tissue with alcohol (povidone iodine or chlorhexidine solutions may interfere with the DNA identification tests).
4. When collecting auricular tissue, ensure that the sampling location does not interfere with animal identification, or if using ear punch as a method of identification that the code is clearly understood. Disinfect ear-punch instrument between animals using the bead sterilizer followed by cooling with saline.
5. When collecting tail tissue, push the skin toward the tip of the tail and using sterile scalpel, or razor blade, cleanly excise the distal 0.5 cm of tail. The blade as well as the surface the tail is placed should be sterile prior to use and between animals. A bead sterilizer can provide complete sterilization and accomplish hemostasis by cauterization. A disposable blade is recommended to prevent cross-contamination when collecting tail tissue for genetic analysis.
6. If the proper procedures are followed, the yield of DNA from 0.5 cm of tail should exceed 50 micrograms, enough for multiple analyses. The yield of DNA does not proportionally increase as tail fragments larger than 0.5 cm are used. If small amounts of DNA are required, investigators should consider taking only 0.2 cm of tail.
7. For more detailed information on tissue collection techniques a PowerPoint presentation entitled ***Tail Snips, Tattoos and Identification Procedures -Training Presentation*** can be viewed at:
http://www3.research.usf.edu/cm/docs/powerpoint/Tails_n_Tatts_Training_files/frame.htm
8. Place tissues into a labeled micro-centrifuge tube, and handle according to PI's instructions.
9. Following tissue collection ensure hemostasis using digital pressure, silver nitrate sticks, Clotisol, or a styptic powder with benzocaine.
10. Ensure that the animal has sufficiently recovered from anesthesia prior to returning it to the housing room.
11. Check tail daily for 7 days to ensure tip is healing.

Approved:

Date: